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EFFECT OF HYPEROXIA ON MAXIMAL OXYGEN UPTAKE, BLOOD ACID-BASE BALANCE, AND LIMITATIONS TO EXERCISE TOLERANCE

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ABSTRACT

EFFECT OF HYPEROXIA ON MAXIMAL OXYGEN UPTAKE, BLOOD ACID-BASE BALANCE, AND LIMITATIONS TO EXERCISE TOLERANCE. **Todd A. Astorino And Robert A. Robergs**. **JEPonline**. 2003;6(2):8-20. Hyperoxia, or an increase in inspired oxygen concentration, has been used by scientists to examine exercise metabolism and physical work capacity. It is apparent that hyperoxia increases VO₂max and exercise tolerance due to an increase in O₂ supply to contracting muscle. Furthermore, hyperoxia increases P_aO₂, which may promote an enhanced diffusion of O₂ in skeletal muscle. Compared to normoxia, hyperoxia may reduce PCr degradation during the metabolic transient, attenuating the magnitude of cellular disturbance characteristic of near-maximal to maximal exercise. These aforementioned increases in exercise tolerance during hyperoxia are not due to alterations in ventilation, lactate (La⁻), or acid/base balance in hyperoxia, as previous data report no change in these parameters compared to normoxia. In addition, it is recommended that researchers take special precautions to ensure the accuracy of gas exchange data in hyperoxia.

Key Words: VO₂max, central limitation, S_aO₂, O₂ breathing, lactate

INTRODUCTION

Hyperoxia is defined as an increase in the inspired oxygen (O₂) concentration. Hyperoxia can be administered via autologous blood reinfusion (1,2), breathing oxygen-rich air (fractional inspired oxygen content (F_IO_2) > (0.2093) (3-7), and exposure to hyperbaria (arterial partial pressure of oxygen (P_aO_2) ~ 500 Torr) (8). During the last two decades, hyperoxia has been widely used by researchers to examine limitations to maximal oxygen uptake (VO₂max), so it is necessary to summarize these findings. In addition, since much additional research has been completed since the most recent review of hyperoxia and exercise tolerance (4), a thorough and more current review of the effects of increased O₂ content on exercise tolerance, cardiovascular function, and acid/base balance is warranted. Search criteria for this review included all studies in which healthy subjects completed incremental exercise to volitional fatigue under hyperoxic conditions.

EFFECT OF HYPEROXIA ON EXERCISE TOLERANCE

Soon after the discovery of O_2 , its effects on exercise capacity were determined. Early anecdotal reports (9-10) suggested that breathing pure O₂ increased exercise tolerance, but shortcomings in research design and laboratory equipment minimized the validity of these data. During the next 30 years, research with improved experimental design and methodology supported early findings showing that hyperoxia improved work tolerance independent of exercise mode. However, psychological effects of breathing hyperoxic gas could not be eliminated as a cause of improved performance since control groups, trial randomization, and masking of subjects were not employed. One of the first well-controlled studies (11) to investigate changes in exercise



Figure 1. Increases in exercise tolerance with increasing F_1O_2 . Adapted from Wilson and Welch. (11).

tolerance in hyperoxia required active men to complete treadmill exercise to exhaustion in room air and four hyperoxic inspired gas fractions (40, 60, 80, and 100 %O₂). Results are shown in Figure 1.

Hyperoxia, VO₂max and Limitations to Exercise

Run time to exhaustion increased in a near-linear fashion from 40 to 100 %O₂. It is interesting to observe that compared to 80 %O₂, run time was still increased in 100 %O₂, contrary to earlier reports (12) that exercise tolerance plateaus at F_1O_2 greater than 0.66. Subsequent work has shown that compared to normoxia, acute administration of hyperoxia enhances exercise capacity during treadmill running (13-15), submaximal cycle ergometry (16-18), and all-out rowing (Table 1) (6-7). The magnitude of the performance benefit varies, however, with the specific variable measured in the study, as it appears that peak workload increases to a lesser degree than time to exhaustion during maximal or supra-maximal exercise. These data suggest that hyperoxia increases the capacity to complete submaximal and/or high-intensity exercise.

Author (yr)	Subjects	% Increase in Ex. Performance	Parameter
Margaria (72)	11 healthy men	19.0	Time at supramaximal
			workload
Fagraeus (73)	11 healthy men	15.1*	Time at supramaximal
			workload
Linnarsson (74)	6 healthy men	20.0*	Peak workload
Davies (74)	5 healthy men	1.0	Peak workload
Adams (80)	6 male runners	26.4*	Time at 90 % VO ₂ max
Buick (80)	11 track athletes	31.0	Time at 95 % VO ₂ max
Wilson (80)	10 healthy men	21.8	Time at 8 mph
Hogan (83)	6 healthy men	5.9	Peak workload
Hogan (84)	6 healthy men	22.0	Time at 90 % VO ₂ max
Powers (89)	7 trained runners	5.3	Peak workload
Plet (92)	11 young men and women	41.0*	Time at 80 % VO ₂ max
Chick (93)	5 healthy men	32.3*	Time at 85 % Wmax
Knight (93)	11 trained cyclists	8.7*	Peak workload
Mateika (94) 8 healthy men		13.0*	Incremental exercise
			time
Peltonen (95)	6 trained rowers	6.5*	Peak workload
Nielsen (98)	11 trained oarsmen	3.2	Peak workoad
Hogan (99)	6 men and women	14.0*	Incremental exercise
			time
Richardson (99)	5 trained cyclists	12.1	Peak workload
Linossier (00)	5 healthy men	45.0*	Maximal exercise time
Harms (01)	25 female runners	57.0*	Time at peak work rate
Peltonen (01)	6 trained men	5.5	Peak workload
Astorino (01)	20 healthy men	7.4*	Peak workload

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* = p < 0.05

EFFECT OF HYPEROXIA ON VO₂MAX

An initial explanation for this enhanced performance in hyperoxia is a greater VO₂max mediated by enhanced oxygen delivery. The Fick Equation proposes that any observed increase in VO₂max is partly attributed to central cardiovascular function (cardiac output) and/or O₂ extraction (arteriovenous O₂ difference). For example, a simple calculation emphasizes the expected change in VO₂max for a hyperoxia-mediated increase in O₂ delivery due to an increase in arterial oxygen content (C_aO₂). Using previously reported values for cardiac output (Q) and arteriovenous O₂ difference (a-vO₂ Δ) in normoxia and 100 %O₂ (5), VO₂max should increase by approximately 11 % (4,547.5 mL/min vs. 4,102.6 mL/min) in hyperoxia. Table 2 shows the magnitude of increase in VO₂max in response to various levels of hyperoxia administered in previous investigations. Recent data from our laboratory (19) revealed that mean VO₂max is 12 % higher in 25 %O₂ (4.34±0.75 L/min) versus normoxia (3.87±0.58 L/min) in moderately-trained young men. This increase is comparable to data reported in

previous research (7,14,17,20) using similar subject populations. So, what factors are responsible for the increase in VO₂max in hyperoxia?

Table 2. Change in VO_2 max, Maximal S_aO_2 , and HKmax reported in Hyperoxia						
Author (yr)	Subjects	Ex.	F_IO_2	∆VO 2max (%)	$\Delta S_a O_2$ (%)	AHRmax
	Ŭ	Mode		2 ()	u 2 ()	(b.min ⁻¹)
Margaria (72)	11 men	TM^1	1.0	+8.1	N/R	+0.8
Ekblom (75)	9 men	TM/CE^2	0.50	+12.6*	+4.0	+2.0
Buick (80)	11 runners	TM	Blood	+5.1*	N/R	N/R
			reinfusion			
Thomson (82)	4 untrained men	TM	Blood	+11.2*	-0.7	+1.0
			reinfusion			
Byrnes (84)	6 men	CE	0.70	+13.0*	N/R	0.0
Spriet (86)	4 runners	TM	Blood	+6.8*	N/R	-12.0
			reinfusion			
Powers (89)	7 runners	CE	0.26	+6.6*	+5.3*	+1.0
Plet (92)	6 men, 5 women	CE	0.55	+3.7, +11.4*	N/R	+2.0, +5.0*
Knight (93)	12 cyclists	CE	1.0	+8.1*	+3.7	+0.4
Peltonen (95)	6 rowers	RE^{3}	0.62	+11.1*	N/R	+11.0
Cardus (98)	6 men and	CE	1.0	+16.4	N/R	+2.0
	women					
Nielsen (98)	11 oarsmen	RE	0.30	+13.3*	+5.4	-4.0
Richardson (99)	5 cyclists	KE^4	1.0	+18.5*	+1.5	-4.0
Astorino(01)	20 healthy men	CE	0.25	+12.1*	+3.1	+1.9
Harms (01)	25 trained	TM	0.26	+6.3*	+5.2*	+1.0
	women					
Peltonen(01)	6 trained men	CE	0.32	+14.0*	N/R	N/R

1 able 2. Change in VO ₂ max, Maximal S_3O_2 , and HKmax reported in Hyperox	Table 2.
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¹ = treadmill, ² = cycle ergometer, ³ = rowing ergometer, ⁴ = one-leg knee extension, N/R = not reported, * = significant increase (p<0.05) in VO₂max, S_aO₂, and HR compared to normoxia

First, an enhanced O_2 delivery (mediated by increases in O and/or C_2O_2) may explain increases in VO₂max coincident with hyperoxia. In response to reinfusion of 2 U of blood eliciting a 11.2 % increase in VO_2max (2), C_aO_2 was significantly higher (21.9±1.2 mL/dL vs. 19.7±0.7 mL/dL), whereas Q was similar except at VO₂max. In another study, O₂ delivery at VO₂max was 11 % higher in 100 %O₂ (4.18±0.23 L/min) compared to room air $(3.77\pm0.2 \text{ L/min})$, due to an increased C_aO₂ and no change in Qlegs (5). The near-linear increase in VO₂max in response to increasing O₂ delivery in hyperoxia from previous studies is shown in Figures 2a-c.

The hyperoxia-mediated increase in C_aO_2 is due to marked increases in $S_aO_2(3,7,19)$ and to a lesser extent, hemoglobin concentration ([Hb]) (1,14). In fact, pioneering data from Powers et al. (1) demonstrated that acute hyperoxia (26.2 %O₂) eliminates appearance of arterial hypoxemia in highly-trained runners, resulting in a significantly higher VO₂max compared to room air (74.7±1.2 vs. 70.1±1.9 mL/kg/min). Trained runners who did not desaturate (marked decline in S_aO₂) at VO₂max showed no increase in VO₂max while breathing hyperoxic gas mixtures. Consequently, it is evident that hyperoxia enhances VO₂max due to an increase in oxygen delivery to active muscle.



Figure 2. Relationship between VO₂max and O₂ delivery in previous research; (a) in response to blood reinfusion (1), (b) during rowing ergometry in normoxia and 30 %O₂ (7), (c) during cycle ergometry in normoxia and 100 %O₂ (5), and (d) during single-leg knee extension in normoxia and 100 %O₂ (32).

Does this hyperoxia-driven increase in O_2 delivery allow athletes to overcome central limitations to VO₂max? The contention that central cardiovascular function regulates VO₂max is not new, as this theory was originally introduced in classic research from A. V. Hill's laboratory (21,22). During this research, subjects ran on an 85 m grass track at various speeds for three minutes while gas exchange was obtained every 30 s with Douglas bags. Hill and colleagues concluded that a running speed of 16 km/hr and VO₂max equal to 4 L/min represented the maximal athletic performance in man. To explain the leveling off in oxygen uptake at these high running speeds, Hill commented (p. 156)..."In running, the oxygen requirement increases continuously as the speed increases, attaining enormous values at the highest speeds: the actual oxygen intake, however, reaches a maximum beyond which no effort can drive it. The oxygen intake may attain its maximum and remain constant merely because it cannot go any higher owing to the limitations of the circulatory and respiratory system."

This brief synopsis of data collected almost 80 years ago represents the initial interpretation of a VO₂ plateau at VO₂max to represent a central limitation to VO₂max. That is to say, at VO₂max, oxygen delivery to the working muscle is interpreted to be inadequate to meet the ever-increasing oxygen demand. Subsequent research (23-25) examining the VO₂ response to discontinuous incremental exercise corroborated the paradigm developed by Hill that O₂ delivery limits VO₂max. Two decades later, in response to 43 % of adult men exhibiting a plateau in VO₂ at VO₂max, Cumming and Borysyk (26) commented (p. 20) that..."To some extent, they (criteria to confirm incidence of VO_2max) are all indicators that the oxygen transport system is fully taxed and exercise is increasingly carried out anaerobically." Similarly, Shephard (27) noted (p. 759)..."During treadmill exercise, maximum effort is halted by central circulatory failure." In addition, it was reported that in men exercising to volitional fatigue, VO_2 max and O_2 delivery are markedly attenuated in 12 % O_2 and 15 %O₂ compared to normoxia (28). Consequently, it is apparent that O₂ delivery to contracting muscle partly limits VO_2 max, and increases in O_2 delivery via hyperoxia promote increases in VO₂max and exercise tolerance.

Peripheral Limitations to VO₂max

Limitations to VO₂max may also exist in the periphery, within skeletal muscle. Saltin et al. (29) used a singleleg cycling protocol to examine central and peripheral adaptation to four weeks of different types of training. Results demonstrated that in response to sprint and endurance training, the trained leg expressed significant increases in VO₂max, and significant decrements in blood La⁻ and heart rate at submaximal work rates, compared to the untrained leg. The authors concluded that peripheral factors play as important a role in limiting exercise performance as central cardiovascular function. It is also regarded that substantial muscle La⁻ accumulation during incremental exercise is associated with onset of volitional fatigue. Early reports from Welch's lab (11) stated that enhanced performance with hyperoxia may be due to a decreased anaerobiosis and a subsequent increase in oxidative metabolism. An enhanced carbon dioxide production (VCO₂) may represent greater oxidative ATP production, yet few studies (7,30) have demonstrated significant increases in VCO₂ in hyperoxia. Therefore, it is necessary to examine changes in VCO₂, ventilation, and blood La⁻ reported in previous research.

Differences in VCO₂, ventilation, and blood La⁻ from previous studies comparing exercise tolerance in normoxia and hyperoxia are shown in Table 3. Recent data (7) revealed a significantly higher VCO₂ at VO₂max in 30 %O₂ in trained rowers completing incremental rowing, although the unique mode of exercise may explain this finding. In addition, VCO₂ was higher during submaximal cycle ergometry in 55 %O₂ (30), although this result was observed in only ten subjects. Data from our laboratory in 20 healthy men (19) demonstrated no difference in maximal or submaximal VCO₂ when gas fractions ranging from 25-35 %O₂ are inspired, although VCO₂ was higher (p>0.05) at VO₂max in hyperoxia. This makes sense, as a greater maximal power output in hyperoxia (19, 31-33) should foster a greater VCO₂ at VO₂max. It has been reported that hyperoxia greater than 60 %O₂ abolishes peripheral chemoreceptor activity (11), thus blunting the ventilatory response. Nevertheless, the majority of previous research (3,5,14,19) reports similar ventilation at VO₂max in hyperoxia and normoxia. If maximal ventilation is similar, non-metabolic CO₂ production may also be similar, leading to no change in VCO₂ at VO₂max in hyperoxia.

Author (yr)	VCO ₂ (L/min)	Ventilation (L/min)	Blood La ⁻ (mmol/L)		
Fagraeus (73)	$4.5(0.2), 4.4(0.3)^{1,2}$	117.6(6.5),134.1(7.5)	14.5(0.6),14.6(0.8)		
Ekblom (75)	N/R^3	139.9(5.4),157.3(7.3)	N/R		
Welch (77)	3.1, 3.4	88.8, 107.6	6.1, 8.1		
Buick (80)	N/R	78.9, 80.2	1.8, 2.2		
Thomson (82)	N/R	137.6(31.4),129.3(29.2)	6.7(0.8),6.1(0.9)		
Byrnes (84)	ND^4	109.2(19.9),115.6(5.0)	9.4(3.5), 9.9(2.6)		
Spriet (86)	N/R	N/R	4.7(0.3),12.2(1.7)		
Powers (89)	5.8(0.4), 5.4(0.2)	129.7(4.4),126.1(4.7)	N/R		
Plet (92)	N/R	141.9(5.2),157.3(7.9)	9.3(0.9),10.6(1.1)		
Knight (93)	N/R	161.7(4.1),161.4(4.8)	8.5(0.4), 9.5(0.5)		
Peltonen (95)	N/R	174.5(18.0),181.8(19.1)	13.2(3.6),13.7(5.4)		
Cardus (98)	N/R	95.0(12.0),110.0(6.0)	9.8(0.6), 10.5(0.4)		

Table 3. Alterations in Maximal VCO₂, Ventilation, and Blood La⁻ in Hyperoxia and Normoxia

Central Nervous System Limitations to VO₂max

Previous research demonstrated alterations in catecholamine release with manipulation of F_IO_2 . Augmented sympathetic activation has been identified as a mechanism to increase HR and blood flow at a given work rate in acute hypoxia (34). In response to submaximal cycle ergometry in room air and 100 %O₂, a significant reduction in norepinephrine and epinephrine levels was shown after 10 and 15 min of exercise (35). However, during prolonged cycle ergometry at 67 %VO₂max (36), no difference (p>0.05) in catecholamine concentration was evident between room air and 60 %O₂, leading the authors to conclude that changes in metabolic or cardiorespiratory function were independent of catecholamines. Consequently, it is unlikely that sympathetic activity alters exercise tolerance in hyperoxia.

It is also plausible that enhanced motor unit recruitment may explain increased exercise tolerance in acute hyperoxia. A recent study (34) required six healthy men to perform forearm exercise and cycle ergometry to exhaustion at sea level and after one month of high altitude (5,050 m) acclimatization. During the altitude trial, $100 \% O_2$ was administered to subjects prior to fatigue, and exercise ensued for an additional 3 min. During all trials, electrodes were placed on the right vastus lateralis to acquire electromyographic (EMG) data. Results

showed that integrated EMG significantly increased throughout the additional 3 min of exercise breathing 100 $\%O_2$, suggesting greater recruitment of inactive fibers during this bout. Nevertheless, these data were obtained in only six subjects, and the additional influence of chronic hypoxia on motor unit recruitment causes these data to be speculative. During incremental treadmill running in healthy men, Mateika et al. (37) reported no differences in root mean square EMG voltage in hypoxia, normoxia, and hyperoxia (66 $\%O_2$). It would be interesting to acquire EMG data from a large number of trained cyclists during incremental exercise in normoxia and hyperoxia to determine if motor unit recruitment is indeed greater under conditions of increased PO₂. This would allow researchers to identify another plausible limitation to exercise tolerance.

EFFECT OF HYPEROXIA ON OXIDATIVE METABOLISM

Past research regarding VCO_2 and ventilation (V_E) does not reveal whether oxidative metabolism is augmented in acute hyperoxia. Early work (38) indicated that glycogen depletion is similar during maximal exercise in normobaria and hyperbaria, a finding supported by a similar rate of glycogenolysis in normoxia and 60 %O₂ (39). However, no change in glycogen breakdown in hyperoxia only infers that aerobic metabolism is similar under conditions of higher inspired PO₂. Eloquent research in *in situ* dog muscle (40) elucidated changes in mitochondrial redox state in normoxia and 100 %O₂. At rest, the cytoplasmic NAD⁺/NADH, estimated from lactate/pyruvate, was in a more oxidized state; whereas, during electrical stimulation no differences were observed between normoxia and hyperoxia. In addition, the mitochondrial redox potential, estimated from enzyme activities of the glutamate dehydrogenase system, was not different both at rest and during stimulation. The finding of a more oxidized redox potential in hyperoxia suggests a greater glycolytic rate at the initiation of exercise. This would not only foster greater pyruvate production and resultant flux through the citric acid cycle, but would also provide additional NADH for the electron transport chain. In contrast, work in humans completing maximal exercise in room air and 60 $\%O_2(41)$ revealed an improved maintenance of concentrations of ATP, ADP, and NADH relative to normoxia. This was also associated with reduced accumulation of IMP, La, creatine, and glucose-6-phospate relative to normoxia. This reduced perturbation of cellular homeostasis would promote lesser acidosis in hyperoxia, leading to a better maintenance of contractile function and thus improved exercise tolerance. However, these findings are based upon muscle data from only five subjects, so these results should be accepted with caution. Ultimately, it appears that cellular metabolism is regulated by O₂, and further research with greater statistical power is warranted to better investigate the contention that oxidative metabolism is augmented in hyperoxia.

EFFECT OF HYPEROXIA ON THE BLOOD LACTATE RESPONSE

To date, only two studies (1,17) have demonstrated a significantly lower blood La⁻ at VO₂max in hyperoxia. The former (1) involved the administration of hyperoxia via graded reinfusion of 3 U of blood, while the Plet et al. (17) study required men to complete cycle ergometry in normoxia and 55 %O₂. However, these studies only used four and five subjects, so these data are speculative. In fact, they are in discord with previous data reporting no differences in maximal blood La⁻ in hyperoxia (Table 3). Previous research (31) not only indicated similar arterial La⁻ at VO₂max in normoxia (9.5±5 mmol/L) and 100 %O₂ (8.5±0.4 mmol/L), but also documented similar La⁻ release, calculated from femoral venous flow and arteriovenous difference, in normoxia (23.7±4.2 mmol[/]min) and hyperoxia (20.1±3.3 mmol[/]min). In isolated dog muscle (42), La⁻ production was similar in normoxia (480.0 \pm 110.0 umol/g) and 100 %O₂ (390.0 \pm 60.0 umol/g). Data from our laboratory (19) in 20 men reveal that maximal blood La⁻ obtained from a heated dorsal hand vein is not different at F_IO₂ equal to 0.25 relative to normoxia (Table 3). However, a trend (p>0.05) was shown for higher blood La⁻ in 30 and 35 $%O_2$, which can be explained by the significantly higher power output in hyperoxia (347.7 \pm 57.6 and 349.2±62.8 Watts, respectively) versus normoxia (325.7±50.8 Watts). This makes sense, since the rate of glycolytic ATP production must be accelerated at the end of incremental exercise to meet the continually increasing ATP demand. This may be due to the greater recruitment of glycolytic, fast twitch motor units (type IIa and IIb) at near-maximal power outputs. Also, results in isolated dog mitochondria (43) demonstrated La

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accumulation in fully aerobic muscle, refuting the claim that La⁻ is not produced in the presence of O_2 . To further elucidate the mechanism by which hyperoxia alters the blood La⁻ response to incremental exercise, future research must address rate of La⁻ clearance and activity of lactate dehydrogenase and pyruvate dehydrogenase in hyperoxia to discern the mechanism by which hyperoxia alters the blood La⁻ response to incremental exercise.

Effect Of Hyperoxia On Blood Acid/Base Balance

Do the lack of differences in ventilation, VCO₂, pH, and La⁻ in hyperoxia represent maintenance of blood acid/base balance under conditions of increased PO₂? Past research examining blood acid/base balance in normoxia and hyperoxia is rather sparse (7,16,31,42). Early work (20) revealed similar values for maximal arterial pH in normoxia (7.23±0.02) and 60 %O₂ (7.22±0.01) in response to maximal treadmill exercise. A similar lack of difference in maximal pH was also demonstrated in elite rowers (7), trained cyclists (31), and healthy, active men (19). However, in response to maximal cycle ergometry, arterialized venous pH in 55 %O₂ (7.39±0.02) was significantly higher versus normoxia (7.35±0.01). Interestingly, in canine gastrocnemius muscle exercising to fatigue (42), arterial hydrogen ion concentration ($[H⁺]_a$) was significantly higher in 100 %O₂ (49.0±1.0 nmol/L) versus normoxia (42.0±2.0 nmol/L). Data from our laboratory (19) using incremental cycle ergometry and simultaneous sampling from a heated dorsal hand vein indicate that maximal pH is not different (p>0.05) in normoxia relative to inspired F_IO_2 equal to 0.25, 0.30, and 0.35 (Figure 3). Consequently, despite a higher maximal power output in hyperoxia, blood pH at VO₂max is similar to normoxia.

So, does the lack of differences in maximal pH in hyperoxia suggest that proton buffering is maintained in hyperoxia? Data from a recent study in our laboratory reveal no differences in maximal arterialized venous bicarbonate concentration ([HCO₃⁻]) at VO₂max in hyperoxia. Similarly, recent research (7) reported comparable arterial [HCO₃⁻] at VO₂max in normoxia (15.0 \pm 1.0 mM) and 30 %O₂ (16.0 \pm 0.0 mM) in trained rowers. In trained men (16), [HCO₃-] was similar at 91 %VO₂max in normoxia and 60 %O₂, although significant differences were revealed between hyperoxia and hypoxia. Hemoglobin also serves as a potent buffer of protons in skeletal muscle. In response to blood reinfusion, arterial hemoglobin concentration ([Hb]) is significantly higher compared to pre-infusion (1,14), yet



Figure 3. Changes in maximal pH decrement with increasing F_1O_2 . Adapted from Astorino (19).

no study to date administering hyperoxic gas fractions has demonstrated a similar effect. Therefore, it is likely that acute hyperoxia does not affect blood acid/base balance, leaving other parameters responsible for enhanced exercise tolerance.

EFFECT OF HYPEROXIA ON PARTIAL PRESSURE OF OXYGEN

An increased arterial partial pressure of oxygen (P_aO_2) is a common result of acute hyperoxia. Several-fold increases in P_aO_2 with hyperoxia have been observed in trained rowers (7), elite cyclists (5,32), elite runners (3), healthy men (17,20,) and in dog muscle (42). In twenty recreationally active men, arterialized venous PO₂ from a heated dorsal hand vein was significantly higher in 25, 30, and 35 %O₂ relative to normoxia (19). These data are shown in Figure 4a. In contrast, maximal P_aO_2 is not enhanced in response to blood reinfusion (1,14). So, does hyperoxia-mediated enhanced PO₂ remove peripheral limitations to VO₂max? Fick's Law of Diffusion states that oxygen uptake is equal to the product of a generalized diffusion conductance (DO₂) and the difference in partial pressure between the red blood cell ($P_{cap}O_2$) and muscle mitochondria ($P_{mt}O_2$). So, it is evident that a direct relationship exists between PO₂ and VO₂max, with alterations in PO₂ via acute hyperoxia causing a commensurate change in VO₂max.



Figure 4a-4d. Relationships between VO2max and PO2 reported in previous literature

Eloquent work from Wagner's lab (32) using maximal single leg knee extension demonstrated significantly higher quadriceps VO₂ in 100 %O₂ (1.28±0.2 L/min) compared to normoxia (1.08±0.2 L/min), which was explained by significant increases in $P_{cap}O_2$ and myoglobin-associated PO₂ ($P_{Mb}O_2$). DO₂ was similar in hyperoxia (34.8±6.7 mmHg) and normoxia (38.1±8.5 mmHg). Figures 4b - 4d demonstrate a linear relationship between previously reported values for VO₂max and $P_aO_2/P_{cap}O_2$ in normoxia and hyperoxia. Overall, these data suggest that in hyperoxia, a greater gradient for diffusion of O₂ from the capillary to the muscle mitochondria enhances VO₂max.

EFFECT OF HYPEROXIA ON HIGH-ENERGY PHOSPHATES

Other peripheral factors involved in regulation of exercise tolerance include phosphocreatine (PCr) and inorganic phosphate (Pi). It is believed that fatigue incurred by shortterm, intense exercise is due to depletion of PCr and resultant increases in Pi that impair the contractile apparatus (44). Concomitant with depletion of PCr is marked cellular perturbation resulting in La⁻ accumulation, glycogen degradation, and impending metabolic acidosis, leading to cessation of activity. Early work (38) demonstrated a trend toward greater depletion of PCr during submaximal exercise in normobaria versus hyperbaria; however, no differences were observed during maximal work. More recent investigation (45) using the submaximal plantar flexion exercise model revealed that muscle [PCr] is better maintained in 100 %O₂ compared to normoxia and hypoxia. This suggests a lower rate of PCr degradation in response to increased PO₂. From muscle biopsy data in five healthy, active men (41), a smaller decrement in muscle [PCr] in response to maximal cycle ergometry in 60 %O₂ (48 mmol/kg dm) versus normoxia (71 mmol/kg dm) was reported. Furthermore, ΔPi was lower in hyperoxia $(41.6\pm11.3 \text{ mmol/kg dm})$ compared to normoxia $(62.5\pm2.4 \text{ mmol/kg dm})$ mmol/kg dm), and [ATP] was preserved in hyperoxia $(22.9\pm0.6 \text{ to } 21.7\pm1.2 \text{ mmol/kg dm})$ compared to a significant decrement (p < 0.05) in normoxia (22.9 \pm 0.5 to 19.7±0.8 mmol/kg dm). Taken together, these data suggest that PCr hydrolysis may be attenuated in acute hyperoxia, resulting in a lower metabolic disturbance and thus enhanced exercise tolerance.

ACCURACY OF GAS EXCHANGE DATA IN HYPEROXIA

Previous research (46,47) questioned the assumption that nitrogen is physiologically inert in hyperoxia, leading to erroneous values for VO₂. To investigate this, Welch and Pedersen (48) compared Douglas bag estimates of VO₂ to those from the Fick Equation and a third equation (VO₂ = V₁ - V_E - VCO₂) in normoxia and 60 %O₂. Results showed that compared to normoxia, the Douglas bag method overestimated VO₂ in hyperoxia, whereas the introduction of a mixing chamber resulted in no differences in VO₂ or VCO₂



Figure 5. Changes in a) VO_2max in response to graded hyperoxia, and VO_2max versus the change in b) power output and c) S_aO_2 in response to graded hyperoxia.

among the three methods. The authors also recommended that specific precautions, including 10 min equilibration with the hyperoxic gas mixture combined with light exercise, be initiated to ensure that net nitrogen exchange is zero. In our laboratory, these guidelines were followed during research designed to investigate the magnitude of increase in VO₂max in response to 25, 30, and 35 %O₂. Our data (19) are

presented in Figure 5a-c. Compared to normoxia, VO₂max was 12, 22, and 38 % higher with graded hyperoxia. However, the rate of change in VO₂ during incremental exercise in 30 and 35 %O₂ (~ 64 ml O₂/Watt/min) was markedly greater than the accepted value of 9-11 ml O₂/Watt/min. Our data demonstrating an overestimation of VO₂max in 30 and 35 %O₂ are shown in Figures 5b and 5c, indicating a relatively minor increase in both maximal power output and S_aO₂ fostered a dramatic increase in VO₂max in hyperoxia. During all testing, gas analyzers showed zero drift after each trial, and gas exchange data were consistently not different after calibration with 35 %O₂ relative to calibration with room air. However, an unidentified source of error led to this overestimation of VO₂max in 30 and 35 %O₂. It is our recommendation that gas exchange indirect calorimetry may not yield precise estimates of VO₂ in hyperoxia, and cardiac output and arteriovenous difference values should be used to determine oxygen uptake in hyperoxia.

SUMMARY

Hyperoxia has been widely used to examine changes in exercise performance, maximal cardiorespiratory capacity, and blood acid/base balance. It is now apparent that hyperoxia enhances exercise tolerance by partially removing central and peripheral limitations to exercise via increases in C_aO_2 and thus O_2 delivery, and P_aO_2 . However, it remains to be clarified whether VO_2 max in hyperoxia is limited by the central nervous system. Furthermore, it has been shown that hyperoxia promotes greater oxidative ATP provision during maximal exercise, although additional investigation is warranted to elucidate this contention. In addition, it is plausible that PCr hydrolysis during the metabolic transient is attenuated in hyperoxia, leading to less cellular disturbance at exercise onset. Researchers should take special precautions to confirm the precision of gas exchange data in hyperoxia. Lastly, acute hyperoxia does not alter maximal ventilation, pH, [HCO₃-], or VCO₂, so the increased VO₂max and exercise tolerance in hyperoxia must be due to factors other than improved acid/base balance.

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REFERENCES

1. Spriet L, Gledhill N, Froese A, Wilkes D. Effect of graded erythrocythemia on cardiovascular and metabolic responses to exercise. *J Appl Physiol* 1986;61(5):1942-8.

2. Thomson J, Stone J, Ginsburg A, Hammett P. O_2 transport during exercise following blood reinfusion. *J Appl Physiol* 1982;53(5):1213-9.

3. Powers S, Lawler J, Dempsey J, Dodd S, Landry G. Effects of incomplete pulmonary gas exchange on VO₂max. *J Appl Physiol* 1989; 66(6):2491-5.

4. Welch H. Hyperoxia and human performance: a brief review. Med Sci Sports Exerc 1982;14:253-62.

5. Knight D, Schaffartzik W, Poole D, Hogan M, BeBout D, Wagner P. Effects of hyperoxia on maximal leg O₂ supply and utilization in men. *J Appl Physiol* 1993;75(6):2586-94.

6. Peltonen J, Rantamaki J, Niitymaki S, Sweins K, Viitasolo J, Rusko H. Effects of oxygen fraction in inspired air on rowing performance. *Med Sci Sports Exerc* 1995;27(4):573-9.

7. Nielsen H, Madsen P, Svendsen L, Roach R, Secher N. The influence of P_aO_2 , pH, and S_aO_2 on maximal oxygen uptake. *Acta Physiol Scand* 1998; 164:89-97.

8. Fagraeus L, Hesser C, Linnarsson D. Cardiorespiratory responses to graded exercise at increased ambient air pressures. *Acta Physiol Scand* 1974; 91:259-74.

9. Douglas C, Haldane J. The effects of previous forced breathing and oxygen inhalation on the distress caused by muscular work. *J Physiol (London)* 1909-1910; 39: i-iv.

10. Hill L, Flack M. The influence of oxygen inhalations on muscular work. *J Physiol (London)* 1910;40:347-72.

11. Wilson G, Welch H. Effects of hyperoxic gas mixtures on exercise tolerance in man. *Med Sci Sports* 1975;7:48-52.

12. Bannister R, Cunningham D. The effects on the respiration and performance during exercise of adding oxygen to the inspired air. *J Physiol (London)* 1954;125:118-37.

13. Wilson B, Welch H, Liles J. Effects of hyperoxic gas mixtures on energy metabolism during prolonged work. *J Appl Physiol* 1980;39(2):267-71.

14. Buick F, Gledhill N, Froese AB, Myers E. Effect of induced erythrocythemia on aerobic work capacity. *J Appl Physiol* 1980;48:636-42.

15. Margaria R, Camporesi E, Aghemo P, Sassi G. The effect of O_2 breathing on maximal aerobic power. *Pfleugers Arch* 1972;336:225-35.

16. Adams RP, Welch HG. Oxygen uptake, acid-base status, and performance with varied inspired oxygen fractions. *J Appl Physiol* 1980;49:863-8.

17. Plet J, Pedersen P, Jensen F, Hansen J. Increased working capacity with hyperoxia in humans. *Eur J Appl Physiol* 1992;65:171-7.

18. Chick T, Stark D, Murata G. Hyperoxic training increases work capacity after maximal training at moderate altitude. *Chest* 1993;104:1759-62.

19. Astorino T. Effect of graded hyperoxia on VO_2max and acid-base balance during exercise testing to volitional fatigue. [dissertation]. Albuquerque: University of New Mexico. 2001.

20. Ekblom B, Huot R, Stein EM, Thorstensson A. Effect of changes in arterial oxygen content on circulation and physical performance. *J Appl Physiol* 1975; 39:71-5.

21. Hill AV, Lupton H. Muscular exercise, lactic acid, and the supply and utilization of oxygen. *Q J Med* 1923;16:135-71.

22. Hill AV, Long C, Lupton H. Muscular exercise, lactic acid, and the supply and utilization of oxygen. Part IV. Methods of studying the respiratory gas exchanges in man, during rapid alterations produced by muscular exercise, and while breathing various gas mixtures. *Proc Res Soc London [Biology]* 1924;97:84-96.

23. Taylor H, Buskirk E, Henschel A. Maximal oxygen intake as an objective measure of cardiorespiratory performance. *J Appl Physiol* 1955;8:73-80.

24. Mitchell J, Blomqvist, G. Maximal oxygen uptake. N Engl J Med 1971;284:1018-22.

25. Wyndham C, Strydom N, Maritz J, Morrison J, Peter J, Potgieter Z. Maximal oxygen intake and maximum heart rate during strenuous work. *J Appl Physiol* 1959;14: 927-36.

26. Cumming G, Borysyk L. Criteria for maximal oxygen uptake in men over 40 in a population survey. *Med Sci Sports* 1972;4:18-20.

27. Shephard R. Tests of maximal oxygen intake, a critical review. Sports Med 1984;1:99-124.

28. Roca J, Hogan M, Story S, BeBout D, Haab P, Gonzalez R et al. Evidence for tissue diffusion limitation of VO₂max in normal humans. *J Appl Physiol* 1989;67(1):291-9.

29. Saltin B, Nazar K, Costill D, Stein E, Jansson E, Essen B et al. The nature of the training response; peripheral and central adaptations to one-legged exercise. *Acta Physiol Scand* 1976; 96:289-305.

30. Prieur F, Busso T, Castells J, Bonnefoy R, Benoit H, Geyssant A et al. Validity of oxygen uptake measurements during exercise under moderate hyperoxia. *Med Sci Sports Exerc* 1998;30(6):958-62.

31. Knight D, Poole D, Hogan M, BeBout D, Wagner P. Effect of inspired oxygen concentration on leg lactate release during incremental exercise. *J Appl Physiol* 1996;81(1):246-51.

32. Richardson R, Leigh J, Wagner P, Noyszewski E. Cellular PO₂ as a determinant of maximal

mitochondrial O₂ consumption in trained human skeletal muscle. J Appl Physiol 1999; 87(1): 325-31.

33. Hogan MC, Cox RH, Welch HG. Lactate accumulation during incremental exercise with various inspired oxygen fractions. *J Appl Physiol* 1983;55(4):1134-0.

34. Kayser B, Narici M, Binzoni T, Grassi B, Cerretelli P. Fatigue and exhaustion in chronic hypobaric hypoxia: influence of exercising muscle mass. *J Appl Physiol* 1994;76:634-40.

35. Hesse B, Kanstrup I, Christensen N, Ingemann-Hansen T, Hansen J, Halkjaer J et al. Reduced norepinephrine response to dynamic exercise in human subjects during O₂ breathing. *J Appl Physiol* 1981;51:176-8.

36. Howley E, Cox R, Welch H, Adams R. Effect of hyperoxia on metabolic and catecholamine responses to prolonged exercise. *J Appl Physiol* 1983;54:59-63.

37. Mateika J, Duffin J. The ventilation, lactate, and electromyographic thresholds during incremental exercise tests in normoxia, hypoxia, and hyperoxia. *Eur J Appl Physiol* 1994;69:110-8.

38. Linnarsson D, Karlsson J, Fagraeus L, Saltin B. Muscle metabolites and oxygen deficit with exercise in hypoxia and hyperoxia. *J Appl Physiol* 1974;36:399-402.

39. Graham T, Pedersen P, Saltin B. Muscle and blood ammonia and lactate responses to prolonged exercise with hyperoxia. *J Appl Physiol* 1987;63(4):1457-62.

40. Wolfe B, Graham T, Barclay J. Hyperoxia, mitochondrial redox state, and lactate metabolism of in situ canine muscle. *Amer J Physiol* 1987;253(22):C263-8.

41. Linossier M-T, Dormois D, Arsac L, Denis C, Gay J, Geyssant A et al. Effect of hyperoxia on aerobic and anaerobic performances and muscle metabolism during maximal cycling exercise. *Acta Physiol Scand* 2000;168:403-11.

42. Hogan MC, Welch, HG. Effect of altered arterial oxygen tensions on muscle metabolism in dog skeletal muscle during fatiguing work. *Amer J Phys* 1986;251(20):C216-22.

43. Connett R, Honig C, Gayeski T. Lactate accumulation in fully aerobic, working, dog gracilis muscle. *Amer J Phys* 1984;246:H120-8.

44. MacLaren D, Gibson H, Pary-Billings M. A review of metabolic and physiological factors in fatigue. *Exerc Sports Sci Rev* 1989; 17:229-36.

45. Haseler L, Richardson R, Videen J, Hogan J. Phosphocreatine hydrolysis during submaximal exercise: the effect of F_IO₂. *J Appl Physiol* 1998; 85(4):1457-63.

46. Cissik J, Johnson R, Rokosch D. Production of gaseous nitrogen in human steady-state conditions. *J Appl Physiol* 1972;32(2):155-9.

47. Wilmore J, Costill D. Adequacy of the Haldane transformation in the computation of exercise VO_2 in man. *J Appl Physiol* 1975;35(1):85-9.

48. Welch H, Pedersen P. Measurement of metabolic rate during hyperoxia. J Appl_Physiol 1981;51:725-31.

49. Harms C, McClaran S, Nickele G, Pegelow D, Nelson W, Dempsey J. Effect of exercise-induced arterial O₂ desaturation on VO₂max in women. *Med Sci Sports Exerc* 2000;32:1101-8.

50. Peltonen J, Tikkanen H, Rusko H. Cardiorespiratory responses to exercise in acute hypoxia, hyperoxia, and normoxia. *Eur J Appl Physiol* 2001;85:82-8.